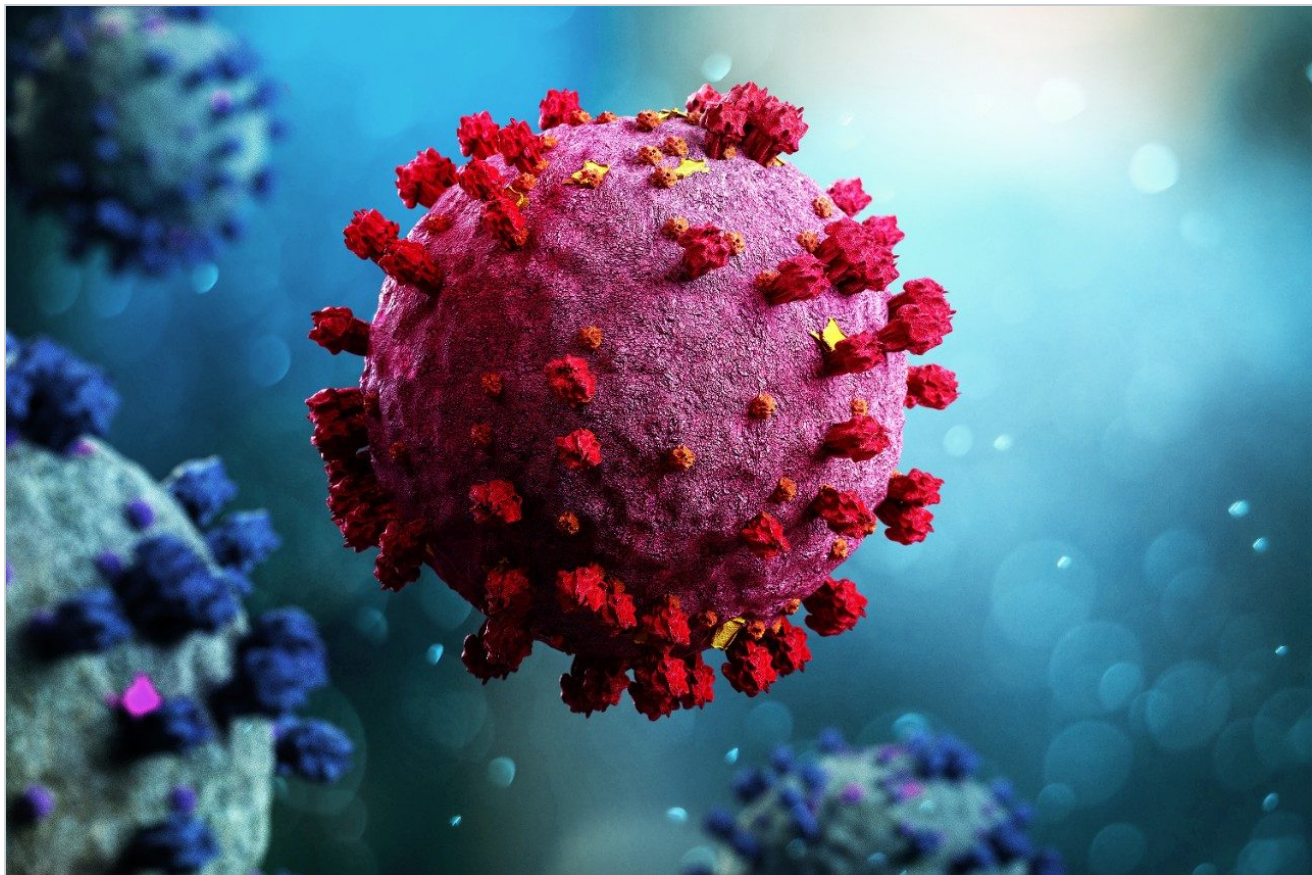


# Comprehending COVID-19: Rapid and Sensitive Characterization of N-Glycans from SARS-CoV-2 Spike Protein

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Xiaoxiao Liu, Matthew A. Lauber

Waters Corporation





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This is an Application Brief and does not contain a detailed Experimental section.

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## Abstract

The global COVID-19 pandemic has resulted in extensive efforts to develop vaccines to the novel coronavirus. Identifying vaccine targets relies on robust analytical methods to understand SARS-CoV-2 structural biology. This work is focused on understanding the N-glycosylation profile of the SARS-CoV-2 spike protein, which has emerged as a potential target for vaccine development. As glycans often dictate critical glycoprotein structure and function, understanding SARS-CoV-2 spike protein glycans is essential to further therapeutic development.<sup>1</sup> This work utilizes the GlycoWorks *Rapi*Fluor-MS N-Glycan Kit to easily and rapidly detect SARS-CoV-2 spike protein N-Glycans. As a result, 42 major glycan peaks were identified, two of which are tentatively assigned as doubly fucosylated. This work motivates further MS/MS analysis to confirm the SARS-CoV-2 spike protein glycosylation profile.

## Benefits

Rapid, sensitive, and easy detection of N-glycans

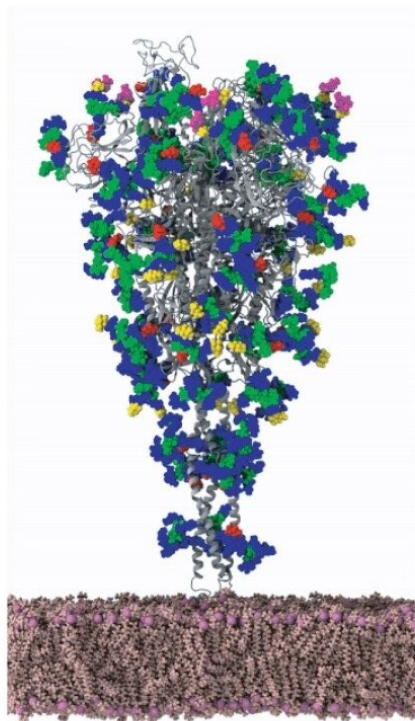
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## Introduction

During the COVID-19 pandemic, scientists across the globe are working to understand SARS-CoV-2 structural biology. Through this work, the SARS-CoV-2 spike protein has been implicated in viral pathogenesis and has thus emerged as a target for vaccine development. Studies show that neutralizing antibodies interact with the spike protein of the novel coronavirus at both peptide and glycan epitopes.<sup>1,2,3</sup> Understanding spike protein glycans is paramount to appropriate therapeutic development as glycosylation can dictate a significant portion of

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the structure, function, conformational dynamics, and drug binding site availability.<sup>1</sup> Therefore, it is critical to characterize glycosylation during the development of new vaccines.



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*Figure 1. The SARS-CoV-2 spike protein (gray) with glycans modeled on its surface.*

*Lorenzo Casalino, Zied Gaieb, and Rommie Amaro, UC San Diego.*

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## Experimental

N-glycans were released, labeled, and purified for hydrophilic interaction chromatography (HILIC) using the GlycoWorks *Rapi*Fluor-MS N-Glycan Kit with optimized DTT reducing conditions for denaturation. HILIC-FLR-MS was performed with an ACQUITY UPLC H-Class Bio System and a Xevo G2-XS QToF Mass Spectrometer.

## LC-MS Conditions

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LC system: ACQUITY UPLC H-Class Bio

Detection: ACQUITY FLR and Xevo G2-XS  
QToF

Vials: QuanRecovery 300 µL

Column(s): ACQUITY UPLC Glycan BEH  
Amide, 1.7 µm, 2.1 x 150 mm

Column temp.: 60 °C

Sample temp.: 8 °C

Injection volume: 1 µL

Flow rate: 0.4 mL/min

Mobile phase A: 50 mM ammonium formate, pH  
4.4

Mobile phase B: Acetonitrile (LC-MS grade)

Gradient: 75–54% Mobile phase B in 35  
minutes

For detailed sample preparation information, please see the GlycoWorks Care and Use Manual. For detailed MS conditions, please see Waters Application Note.

GlycoWorks Care and Use Manual	<a href="https://legacy-stage.waters.com/webassets/cms/support/docs/715004903.pdf">715004903 &lt;https://legacy-stage.waters.com/webassets/cms/support/docs/715004903.pdf&gt;</a>
Waters Application Note	<a href="https://legacy-stage.waters.com/webassets/cms/support/docs/720005850EN.pdf">720005850EN &lt;https://legacy-stage.waters.com/webassets/cms/support/docs/720005850EN.pdf&gt;</a>



## Results and Discussion

42 major glycan peaks were identified (see Figure 2), wherein 2 are tentatively assigned as doubly-fucosylated (see Figure 3). The remaining assignments are: 11 afucosylated glycans, and 29 fucosylated glycans. These glycans can be further grouped into 3 classes, including 6 high mannose glycans, 6 hybrid glycans, 30 complex glycans. These assignments were made based on relative HILIC retention times, glucose unit (GU) values and accurate mass information. Examination by MS/MS analysis and exoglycosidase arrays is warranted in order to confirm identifications.

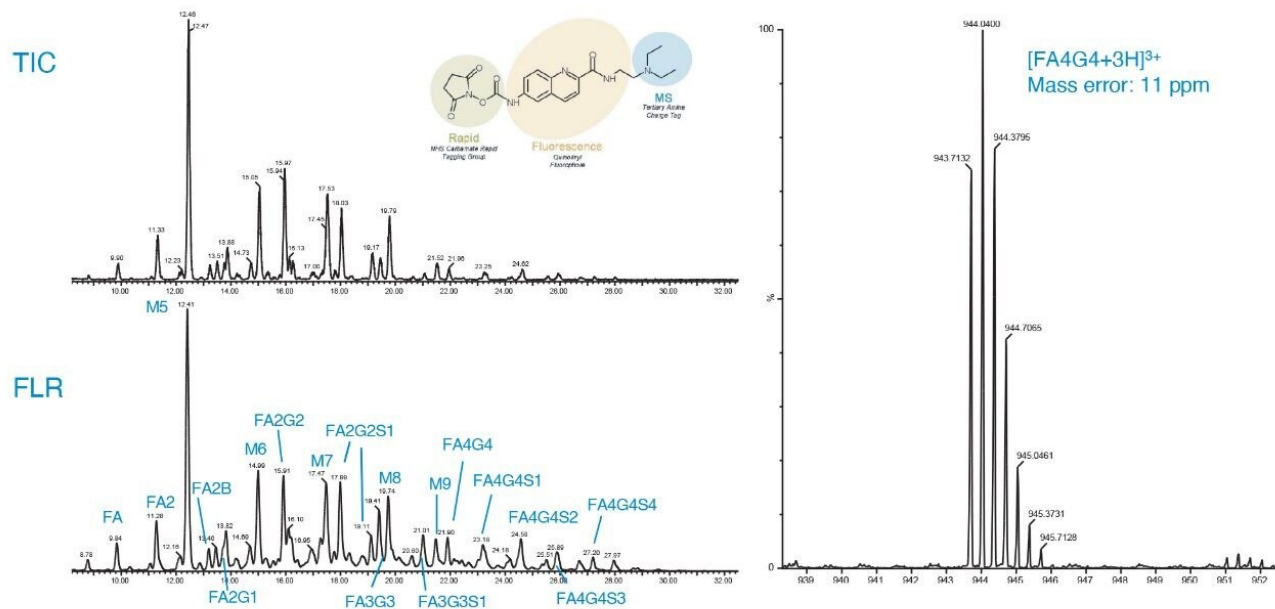


Figure 2. Identification of 42 major glycan peaks by MS.

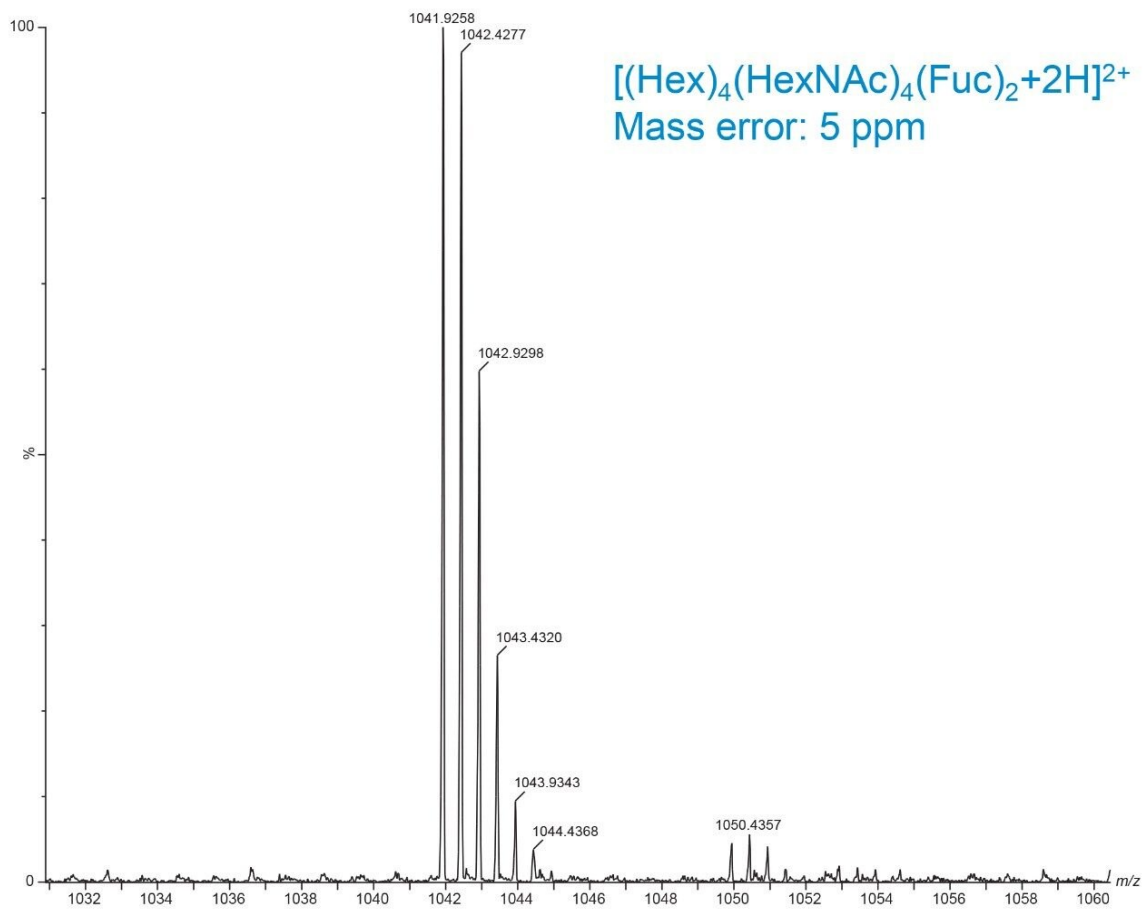


Figure 3. MS spectra of doubly-fucosylated glycans.

## Conclusion

Because the SARS-CoV-2 spike protein is implicated in viral pathogenesis, it has become a target for vaccine development. Efficient therapeutic development relies on a solid structural and functional understanding of the SARS-CoV-2 spike protein target. Understanding the glycan profile is critical to a complete structural and functional understanding of SARS-CoV-2 spike protein. As a result, rapid and accurate glycan analysis is necessary to identify and develop promising new COVID-19 therapies. This work demonstrates the ability to rapidly and easily detect SARS-CoV-2 N-glycans. 42 major glycan peaks were identified. Interestingly, two peaks

were assigned as doubly-fucosylated. This curious finding calls for further corroboration through examination by MS/MS and exoglycosidase arrays.

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## References

1. Novokmet, Mislav *et al.* Understanding Glycans in COVID-19 Drug Design. <https://www.genengnews.com/insights/understanding-glycans-in-covid-19-drug-design/> <  
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  2. Pinto, D. *et al.* Structural and Functional Analysis of a Potent Sarbecovirus Neutralizing Antibody. *bioRxiv* 2020.04.07.023903 (2020). doi: <https://doi.org/10.1101/2020.04.07.023903>
  3. Stawiski, E.W. *et al.* Human ACE2 Receptor Polymorphisms Predict SARS-CoV-2 Susceptibility. *bioRxiv* 2020.04.07.024752 (2020). doi: <https://doi.org/10.1101/2020.04.07.024752>
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