

An Analytically Validated Bioanalytical Clinical Research Method for the Quantification of Acylcarnitines in Urine

Leanne Nye, Nyasha Munjoma, Ian D. Wilson

Waters Corporation

For research use only. Not for use in diagnostic procedures.

Abstract

This application note describes an analytically validated bioanalytical clinical research method for the quantification of low concentration acylcarnitines in urine.

Benefits

- Direct analysis of carnitine and acyl carnitines in urine using gradient reversed-phase UltraPerformance Chromatography (UPLC) combined with tandem quadrupole MS
 - Rapid analysis
 - Minimal sample preparation, without the need for derivatization prior to analysis
-

Introduction

The amino acid derivative carnitine is involved in lipid metabolism, transporting long-chain acyl groups from fatty acids to the mitochondrial matrix, where they are broken down to form usable energy via the citric acid cycle. The analysis of these acylcarnitines in blood, urine, and plasma for clinical research studies is typically performed by tandem quadrupole MS coupled to gas chromatography, high-performance liquid chromatography, or capillary electrophoresis.² In this application note, we describe an analytically validated bioanalytical clinical research method for the quantification of low concentration acylcarnitines in urine.^{3,4}

Experimental

Sample description

Urine samples, stored at -80 °C, were left to thaw at 4 °C. A 30 µL aliquot of urine was centrifuged at 10,300 g for 10 minutes. Then, 20 µL of supernatant from each sample was diluted with 65 µL of water, 5 µL of acetonitrile, and 10 µL of the internal standard mixture. The internal standard mixture consisted of propionyl-L-carnitine-d₃; butyryl-L-carnitine-d₃, hexanoyl-L-carnitine-d₃ and dodecanoyl-L-carnitine-d₃ at 250 ng/mL and isovaleryl-L-carnitine-d₉, octanoyl-L-carnitine-d₃ and decanoyl-L-carnitine-d₃ at 125 ng/mL. Tetradecanoyl-L-carnitine-d₃ and hexadecanoyl-L-carnitine-d₃ were also included at 125 ng/mL for testing of these analytes.

Method conditions

The UPLC-MS analysis was performed on an ACQUITY UPLC binary solvent manager, 2777 sample manager (CTC) and column manager (Waters, Milford, MA, USA) interfaced to a Xevo TQ-S Tandem Quadrupole Mass Spectrometer (Waters, Milford, MA, USA).

LC conditions

LC system:	Open Architecture UPLC System
Detection:	Xevo TQ-S
Vials:	Waters certified vials

Column:	ACQUITY UPLC HSS T3 1.8 μm , 2.1 mm x 150 mm
Column temp.:	45 °C
Sample temp.:	4 °C
Injection volume:	2 μL
Flow rate:	0.5 mL/min
Mobile phase A:	0.1% formic acid
Mobile phase B:	0.1% formic acid in acetonitrile Gradient mode elution

MS conditions

MS system:	Xevo TQ-S
Ionization mode:	Positive ion
Acquisition range:	MRM mode
Capillary voltage:	2.75 kV
Collision energy:	14–26 eV
Cone voltage:	8–48 V

Data management

Chromatography software:	MassLynx Software
--------------------------	-------------------

MS software: MassLynx Software

Informatics: TargetLynx Software

Results and Discussion

The chromatographic separation was developed to provide sufficient resolution of the analytes, with a run time of 10 minutes consistent with medium to high throughput analysis, while maintaining the separation between the two pairs of isomeric species isovaleryl-L-carnitine (isoC5) and valeryl-L-carnitine (C5), and isobutyryl-L-carnitine (isoC4) and butyryl-L-carnitine (C4). Due to their relatively low concentrations in urine, compared to carnitine itself and some other high concentration acylcarnitines, the analytes measured in this method, propionyl-L-carnitine (C3), butyryl-L-carnitine (C4), isovaleryl-L-carnitine (isoC5), valeryl-L-carnitine (C5), hexanoyl-L-carnitine (C6), octanoyl-L-carnitine (C8), decanoyl-L-carnitine (C10) and dodecanoyl L-carnitine (C12) need to be quantified using a method optimized for low concentration acylcarnitines. Dodecanoyl L-carnitine (C12), tetradecanoyl-L-carnitine (C14) and hexadecanoyl-L-carnitine (C16) are too low in concentration in the control urine to be quantified using the sample preparation method for this analysis, however, the internal standards are displayed in Figure 1B for the purposes of demonstrating retention time and could allow for testing of these analytes.

LC-MS

The samples were analyzed using a reversed-phased gradient separation with the methodology optimized for analyte resolution and reproducibility. The analytes were detected and quantified by positive ion MRM with optimal MS conditions determined by the infusion of authentic analyte and stable isotope labelled internal standards to determine the maximum response for each transition. For quantification, one transition from each of these precursor ions was used to calculate unknown concentrations. The choice of quantification transition was determined from the ionization energy of each compound.

Typical chromatograms for the UPLC-MS/MS analysis of the acylcarnitine standards are shown in Figure 1A and 1B. Figure 2 shows a typical chromatogram of a urine sample diluted one part in five. All eight acylcarnitines examined were chromatographically resolved. The separation was shown to

be highly reproducible, with retention time coefficients of variance for the acylcarnitine standards at the ULOQQC of <0.20% across the entire validation run (n=18).

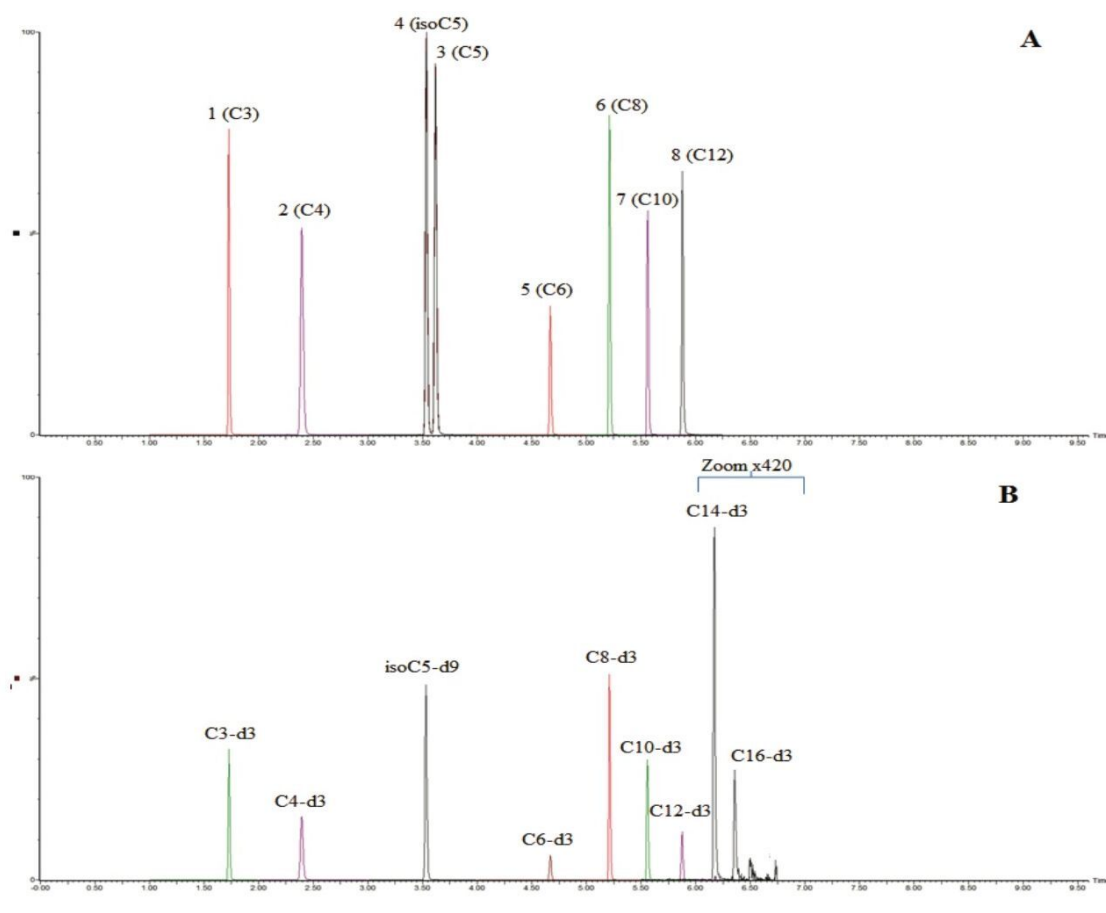


Figure 1. Chromatographic separation of the ULOQQC acylcarnitine standards (A) natural abundance species (B) SIL species. Note at the retention times of C14-d₃ and C16-d₃, B is zoomed in 420 times to show IS peaks

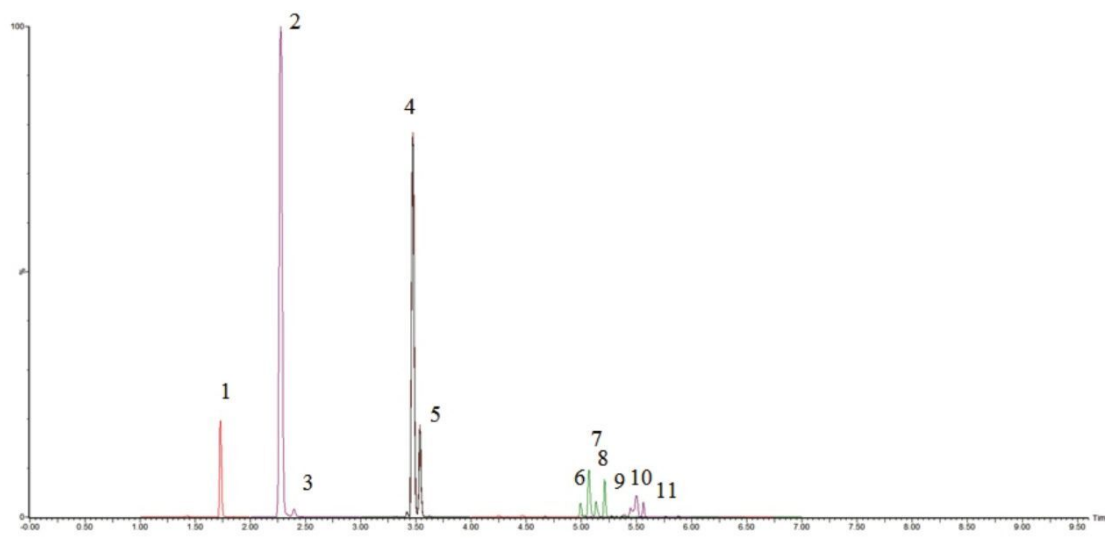


Figure 2. Typical urine sample diluted one part to five. Peak annotation;

- 1) propionyl-L-carnitine,
- 2) isobutyryl-L-carnitine,
- 3) butyryl-L-carnitine,
- 4) isovaleryl L-carnitine,
- 5) valeryl L-carnitine,
- 6, 7, 8) isomers of octanoyl-L-carnitine,
- 9) octanoyl-L-carnitine,
- 10) isomers of decanoyl-L-carnitine,
- 11) decanoyl-L-carnitine

Linearity

The method was linear over the calibration ranges employed for the individual analytes, using a linear fit with a weighting of $1/X$. The correlation coefficient (r^2) values of the individual standard curves were determined to be between 0.988 and 0.999 for all analytes.

Intra-day validation

The method was assessed for accuracy linearity and bias over three days for the concentration ranges 7.5–100 ng/mL for propionyl-L-carnitine (C3); 3–40 ng/mL for butyryl-L-carnitine (C4), valeryl-L-carnitine (C5), isovaleryl-L-carnitine (isoC5), hexanoyl-L-carnitine (C6) and dodecanoyl-L-carnitine (C12); 1.5–20 ng/mL for octanoyl-L-carnitine (C8) and decanoyl-L-carnitine (C10).

All of the analytes monitored showed excellent accuracy and precision. As an example, the coefficient of variance and mean concentration data obtained for the method data for day three are shown in Tables 1 and 2 respectively.

CV	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
Propionyl-L-carnitine, C3	1.67	2.06	2.35	5.88	2.11	4.62
Butyryl-L-carnitine, C4	2.61	2.17	1.89	5.49	1.37	4.12
Valeryl-L-carnitine, C5	1.89	1.46	2.17	5.47	1.30	3.68
Isovaleryl-L-carnitine, isoC5	1.92	2.13	2.12	4.82	1.59	4.25
Hexanoyl-L-carnitine, C6	1.70	1.84	3.02	5.58	1.47	4.76
Octanoyl-L-carnitine, C8	2.58	1.93	2.71	6.39	2.38	4.38
Decanoyl-L-carnitine, C10	3.16	3.89	3.52	8.02	4.31	6.55
Dodecanoyl-L-carnitine, C12	4.50	4.40	10.9	12.10	9.8	14.4

Table 1. Summary of the coefficient of variance for day three

Numbers rounded to three significant figures, or integers, as appropriate.

Mean	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
nominal concentration (ng/mL)	7.5	10	20	30	80	100
Propionyl- L-carnitine C3	7.02	9.42	19.3	29.3	77.6	96.9
nominal concentration (ng/mL)	3	4	8	12	32	40
Butyryl-L-carnitine, C4	2.88	3.85	7.78	11.7	31.1	38.8
Valeryl-L-carnitine, C5	2.73	3.75	7.70	11.6	30.6	38.0
Isovaleryl-L-carnitine, isoC5	2.85	3.83	7.75	11.7	30.8	38.3
Hexanoyl-L-carnitine, C6	3.03	4.08	8.05	11.6	29.9	36.7
Dodecanoyl-L-carnitine C12	3.03	3.80	7.90	11.7	30.0	33.7
nominal concentration (ng/mL)	1.5	2	4	6	16	20
Octanoyl-L-carnitine, C8	1.58	2.12	4.32	6.52	17.1	21.4
Decanoyl-L-carnitine, C10	1.63	2.15	4.40	6.63	17.3	21.6

Table 2. Summary of the mean concentration data for day three

Numbers rounded to three significant figures, or integers, as appropriate.

Inter-day validation

The inter-day validation results are summarized in Table 3 and Table 4. Table 3 summarizes the inter-day coefficients of variance for each QC standard and Table 4 summarizes the inter-day mean concentration for each QC standard. The precision of the back calculated standards across the inter-day validation ranged from 0.89 to 9.75%, while the mean back calculated concentrations obtained for each of the standards gave mean bias values ranging from 3.25 to 8.20%. The Lower Limit of Quantification (LLOQ) was defined as 7.5 ng/mL for propionyl-L-carnitine (C3), 3 ng/mL for butyryl-L-

carnitine (C4), valeryl-L-carnitine, (C5), isovaleryl-L-carnitine (isoC5) and hexanoyl-L-carnitine (C6), and 1.5 ng/mL for octanoyl L-carnitine (C8). However, for dodecanoyl-L-carnitine the LLOQ was 4 ng/mL and decanoyl- L-carnitine (C10) was 2 ng/mL, although the standards were tested to 3 ng/mL and 1.5 ng/mL respectively. The mean bias of the method ranged from -1.7% for decanoyl-L-carnitine, (C10) to +14.7% for dodecanoyl-L-carnitine (C12).

Acylcarnitine	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
Propionyl- L-carnitine, C3	3.61	3.22	2.57	3.57	2.34	3.76
Butyryl-L-carnitine, C4	5.73	6.06	5.10	7.35	6.50	7.03
Valeryl-L-carnitine, C5	3.80	2.52	1.82	3.46	2.35	3.14
Isovaleryl-L-carnitine, isoC5	6.33	5.51	5.64	5.70	4.94	5.15
Hexanoyl-L-carnitine, C6	2.53	2.47	2.49	3.87	2.44	3.92
Octanoyl-L-carnitine, C8	5.41	3.83	5.37	6.53	4.77	5.53
Decanoyl-L-carnitine, C10	7.24	4.74	6.12	7.62	7.37	8.44
Dodecanoyl-L-carnitine, C12	25.2	19.0	8.94	8.56	11.2	12.0

Table 3. Summary of the inter-day coefficient of variance

Numbers rounded to three significant figures, or integers, as appropriate.

Mean	LLOQQC1	LLOQQC2	LQC	MQC	HQC	ULOQQC
nominal concentration (ng/mL)	7.5	10	20	30	80	100
Propionyl- L-carnitine, C3	6.85	9.22	18.7	28.0	74.4	92.7
nominal concentration (ng/mL)	3	4	8	12	32	40
Butyryl-L-carnitine, C4	2.84	3.81	7.62	11.5	30.5	37.9
Valeryl-L-carnitine, C5	2.68	3.64	7.39	11.1	29.4	36.5
Isovaleryl-L-carnitine, isoC5	3.07	4.11	8.25	12.4	32.7	40.6
Hexanoyl-L-carnitine, C6	2.97	3.98	7.96	11.7	30.8	38.2
Dodecanoyl-L-carnitine, C12	3.83	4.71	8.25	12.3	29.8	36.2
nominal concentration (ng/mL)	1.5	2	4	6	16	20
Octanoyl-L-carnitine, C8	1.53	2.04	4.05	6.09	16.2	20.0
Decanoyl-L-carnitine, C10	1.63	2.11	4.11	6.14	15.9	19.8

Table 4. Summary of the inter-day mean concentration data

Numbers rounded to three significant figures, or integers, as appropriate.

Carryover

Carryover was assessed by running a double blank (water, containing no internal standard and no analytes) immediately after a upper limit of quantitation (ULOQ) calibration standard. The results obtained indicate that there was no significant carryover for the acylcarnitine standards. Carryover is the peak area of a compound found in the blank sample calculated as a percentage of the peak

area of the compound in the lower limit of quantification quality control (LLOQQC). The highest percentage carry over at 14.27% was observed for dodecanoyl-L-carnitine. This is within the accepted limit of 20%. The internal standards carryover, at less than 0.98% for all compounds, was negligible.

Conclusion

Based on the use of stable isotope labelled internal standards, a quantitative clinical research method was validated over the range 7.5–100 ng/mL (37.5–500 ng/mL in urine) for propionyl-L-carnitine (C3) and 3–40 ng/mL (15–200 ng/mL in urine) for butyryl-L-carnitine (C4), valeryl-L-carnitine, (C5), isovaleryl-L-carnitine (isoC5) and hexanoyl-L-carnitine (C6). Dodecanoyl-L-carnitine (C12) ranged from 4–40 ng/mL (20–200 ng/mL in urine) while the octanoyl-L-carnitine (C8) range is from 1.5–20 ng/mL (7.5–100 ng/mL in urine) and decanoyl-L-carnitine (C10) from 2–20 ng/mL (10–100 ng/mL in urine). The methodology required only 30 μ L of urine and exhibited excellent analytical sensitivity, robustness, and reproducibility.

The analytical data presented in this application note are intended to demonstrate the robustness of a Waters research method. These data in no way substitute for independent method validation required by any applicable legal or laboratory standards.

References

1. Vernez L, Thormann W, Krähenbühl S (2000) Analysis of carnitine and acylcarnitines in urine by capillary electrophoresis, *J Chromatogr A*, 895 (1–2): 309–16.
2. FDA, Bioanalytical Method Validation. <http://www.fda.gov>. 2001, May.
3. FDA Guidance for Industry Bioanalytical Method Validation. Draft Guidance. September 2013 Biopharmaceutics Revision 1.

Featured Products

[Open Architecture UPLC <https://www.waters.com/10083082>](https://www.waters.com/10083082)

[TargetLynx <https://www.waters.com/513791>](https://www.waters.com/513791)

[Xevo TQ-S <https://www.waters.com/10160596>](https://www.waters.com/10160596)

[MassLynx MS Software <https://www.waters.com/513662>](https://www.waters.com/513662)

720005974, May 2019

©2019 Waters Corporation. All Rights Reserved.